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The synergistic effect between graphene oxide nanocolloids and silicon dioxide nanoparticles for gallic acid sensing

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Abstract

For the first time, the synergistic effect of graphene oxide nanocolloids (nano-GO) and silicon dioxide (silica) nanoparticles (SiO₂-nanoparticles) has been used to modify a glassy carbon electrode (GCE) for the determination of gallic acid (GA). The modified electrode surface was characterised by using scanning electron microscopy (SEM), energy-dispersive X-ray spectroscopy (EDXA) and Fourier transform infrared spectroscopy (FTIR). The electrochemical behaviour of the modified electrode was then studied, using cyclic voltammetry (CV) and differential pulse voltammetry (DPV), showing that the electrode was sensitive to GA in a concentration range of 6.25×10^{-6} to 1.0×10^{-3} mol L⁻¹, with a correlation coefficient R^2 of 0.9956 and a limit of detection of 2.09×10^{-6} mol L⁻¹ (S/N = 3). The proposed method was successfully used for the determination of GA in red wine, white wine and orange juice, with recoveries of 102.3, 95.4 and 97.6%, respectively.

Keywords Graphene oxide nanocolloids · Gallic acid · SiO₂ nanoparticles · Electrochemistry · Cyclic voltammetry · Differential pulse voltammetry

Introduction

Gallic acid (2,3,4-trihydroxybenzoic acid) (GA) is a naturally occurring phenolic compound in plants and is a natural antioxidant with several biological activities. The compound has shown its capacity as a reducing agent in the pharmaceutical industry and also as a homeopathic drug that can be found in tea, red wine, fruits (bananas, blueberries, grapes), beverages and several medicinal plants [1]. In the pharmaceutical industry, it is widely used as a reducing agent in products like Dermatol, Airol and bismuth salts of gallic acid [2]. Several studies had shown GA to have anti-carcinogenic, anti-mutagenic and anti-oxidative properties [3]; hence, it is recommended usage as an antioxidant in human diet. This is because it can reduce the risk of disease development, by the

prevention or slowing down of molecular oxidation in the body [4], known as oxidative stress. Furthermore, oxidative stress has been linked to human diseases like Alzheimer's disease, cardiovascular diseases, Parkinson's disease, diabetes and cancer [5, 6].

With the increasing importance of antioxidants in our diets and our health, especially natural antioxidants like vitamins, carotenoids and phenolic or flavonoid compounds in the reduction of oxidative stress, many in vitro chemically based assays have been developed for the determination of the capacity of these antioxidants. The study of antioxidant properties has been mainly carried out by spectrophotometric methods [7, 8], chromatographic [9, 10], flow injection-chemiluminescence [11] and electrochemical methods [12–14]. However, the main drawbacks to the chromatographic and spectrophotometric methods have been the cost of equipment, coupled with the extensive time-consuming sample preparation procedures as well as the use of expensive and toxic reagents which are detrimental to the environment.

Amongst all the available methods, electrochemical techniques have generated a lot of interest due to their many advantages, including being fast, sensitive, selective, inexpensive and portable, with little or no sample pre-treatment. Furthermore, they are amenable for miniaturisation and do not suffer from colour interference. Many different

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electrochemical methods have been developed in literature for the determination of polyphenols [15–19]. The most common electrochemical techniques for the determination of GA have been CV and differential pulse voltammetry (DPV), which were also used in this work.

Nanomaterials such as SiO₂ and TiO₂ have been previously employed in the modification of carbon paste electrodes [14, 20] for the determination of GA with relatively low limits of detection (LOD) of 2.5×10^{-7} mol L⁻¹ and 9.4×10^{-7} mol L⁻¹, respectively. Although metal oxide nanoparticles with graphite or regular graphene oxide sheet electrodes have been used for gallic acid determination, there has been no attempt in using graphene oxide nanocolloids. Aqueous graphene oxide nanocolloids having high charge density originating from their high edge-to-area ratios are significantly more stable than the regular graphene oxide.

Here in this work and for the first time, the synergetic effect of graphene oxide nanocolloids and SiO₂ nanoparticles has been used to modify a glassy carbon electrode, by a drop-casting method in the determination of the antioxidant capacity of gallic acid. The modified electrode was also used to demonstrate selectivity by the simultaneous determination of gallic acid and uric acid, with a successful determination of GA in orange juice, red wine and white wine.

Experimental

Reagents and materials

Gallic acid (anhydrous, molar mass 170.12 g mol⁻¹), potassium chloride (analytical grade $\geq 99.8\%$), graphene oxide nanocolloids (nano-GO), tetraethyl orthosilicate (TEOS) and aqueous ammonia (28–30%), and potassium hexacyanoferrate (II) trihydrate (analytical grade $\geq 99.95\%$) were purchased from Sigma Aldrich (London, UK). Analytical grade salts of sodium dihydrogen orthophosphate and disodium hydrogen orthophosphate, potassium nitrate, sodium hydroxide and phosphoric acid were from Merck (Darmstadt, Germany). All the reagents unless stated were of the highest purity, with no further purification required. All solutions were freshly prepared with doubly distilled water with a resistance value of 18.2 M Ω cm. All the stock solutions used were protected from light and refrigerated at about 4 °C. Wine samples were commercially available brands (Casillero Del Diablo Cabernet Sauvignon and Sauvignon Blanc) and the orange juice (Innocent Smooth Orange Juice) from ASDA (UK).

Apparatus and methods

All the electrochemical measurements were carried out using an Ivium vertex one potentiostat-galvanostat and analysed

using Iviumsoft software (Eindhoven, Netherlands). A standard three-electrode system was employed to perform all the electrochemical experiments. The cell consisted of a commercially available glassy carbon working electrode with a diameter of $\varnothing = 0.5$ cm, a KCl saturated Ag/AgCl reference electrode and a platinum counter electrode, purchased from BASi (West Lafayette, USA). The 0.1-mol L⁻¹ phosphate buffer solution (PBS) was prepared by mixing the appropriate volumes of 0.2 mol L⁻¹ Na₂HPO₄ and 0.2 mol L⁻¹ NaH₂PO₄ to 50 mL, then diluted to 100 mL with deionised water. Meanwhile, the measurement of the pH conditions was carried out with a Fisher Scientific (Loughborough, UK) Mettler Toledo Benchtop pH meter. The phosphate buffer solutions with pH values 2, 4, 6 and 8 were prepared by adjusting the ratio of Na₂HPO₄ and NaH₂PO₄, whilst using 1×10^{-2} mol L⁻¹ phosphoric acid to reduce the pH and 0.1 mol L⁻¹ sodium hydroxide to increase the pH. Ultrasonication of the polished electrode was done by using a Fisher Scientific ultrasonication bath from Fisher Scientific (Loughborough, UK). All the potentials quoted in the experiments were relative to an Ag/AgCl reference electrode. Statistical analysis of all data was carried out with MS Excel and all the voltammograms and graphs produced using Origin Pro 8 software.

Synthesis of SiO₂ nanoparticles

The synthesis of SiO₂ nanoparticles was carried out according to Venkathathri [21], where aqueous ammonia (3.14 mL) was added to a solution of ethanol (74 mL) and deionised water (10 mL). Then, tetraethyl orthosilicate (TEOS) (6 mL) was added to the previously prepared solution with vigorous stirring at room temperature. This is because sol-gel synthesis is performed at low temperatures, typically at room temperature. The reaction mixture was stirred for 1 h, to yield uniform silica spheres in a sol. The latter was centrifuged, and the residue was washed with distilled water and ethanol three times followed by drying at room temperature to yield the SiO₂ nanoparticles.

Preparation of the electrodes

The synthesised SiO₂ nanoparticles were dispersed in a solution of graphene oxide nanocolloids at a concentration of 1 g/L, being the amount of the nanoparticles with the best result. Then, the mixture was sonicated for about 1 h, to provide a uniform dispersion of SiO₂ nanoparticles in the GO nanocolloids solution. Prior to each experiment, the GCE was polished on a felt lapping pad purchased from Alvatek (Tetbury, UK), with 0.1, 0.3 and 0.05 μ m alumina paste to a mirror finish. Subsequently, the electrode was rinsed with water, and finally, an ultrasonic treatment in water and ethanol was applied, respectively, to remove any particles of alumina slurry on the electrode.

A mixture of graphene oxide (2 mg/mL) solution and SiO₂ nanoparticles (1 mg/mL) in deionised water was prepared by sonicating both compounds for 1 h. After that, the GCE was modified with the prepared solution by drop casting with 5 μ L of the suspended nano-graphene oxide-SiO₂ nanoparticles directly on to the surface of the electrode. The electrode, modified with GCE-nano-graphene oxide-SiO₂ nanoparticles, was then left to dry at room temperature. GCE-SiO₂ and GCE-GO were prepared by drop casting SiO₂ and GO, respectively, on the surface of the GCE. The electrolytes used for the experiment were thoroughly deoxygenated by bubbling/purging with high purity nitrogen gas prior to each experiment.

Electrochemical measurements

Electrochemical analysis of GA at the surface of the nano-GO-SiO₂ nanoparticle-modified GCE was carried out in 0.1 mol L⁻¹ pH 2.0 phosphate buffer solution. The CV was scanned within a range of 0.0 V to + 1.8 V at a scan rate of 100 mVs⁻¹. The differential pulse voltammograms were recorded in a potential range between 0.0 and + 1.2 V with a pulse amplitude of 0.08 V and pulse period of 0.2 s.

CV was used to study 1×10^{-3} mol L⁻¹ potassium ferrocyanide [Fe(CN)₆]³⁻ on the surface of the electrodes, swept over a potential range of - 0.6 to 0.6 V at different scan rates. The voltammograms recorded were used for the determination of the electroactive areas of nano-GO-SiO₂ nanoparticle GCE, nano-GO-GCE and the bare GCE electrode.

Electrochemical impedance spectroscopy was used as a technique to provide information on the impedance changes on the bare and modified GCE electrode surfaces and the Nyquist plots were recorded. The EIS measurement was performed in the frequency range of 100 kHz–0.01 Hz (50 points within the frequency range) and a potential of 0.4 V in 0.1 mol L⁻¹ KCl containing 5 mmol L⁻¹ [Fe(CN)₆]^{3-/4-} redox solution.

Adsorption time of GA on to the modified electrode was studied by DPV measurement and the voltammograms for the determination 1×10^{-3} mol L⁻¹ GA, recorded every minute for 6 min.

The selectivity of the modified electrode was investigated by mixing 5 mL of a 0.48 mmol L⁻¹ GA and 5 mL of a 0.36 mmol L⁻¹ uric acid in 0.1 mol L⁻¹ phosphate buffer solution of pH 2.0 and then cyclic voltammograms are recorded at room temperature, with a scan rate of 100 mVs⁻¹.

The reproducibility and repeatability of the method were tested by measuring eight replicates of 10 mL, 1×10^{-3} mol L⁻¹ GA in a 0.1 mol L⁻¹ phosphate buffer of pH 2.0, using three different nano-GO-SiO₂ nanoparticle-modified GCEs, and the DPVs are recorded.

The interference of foreign ions in the determination of GA was investigated, by adding 5 mL of 1×10^{-3} mol L⁻¹ solution of GA into 5 mL of 0.1 mol L⁻¹ solutions containing K⁺, Ca²⁺, Fe³⁺ and Na⁺ ions, whilst 5 mL of 1×10^{-3} mol L⁻¹ solution of

ascorbic acid, caffeine and caffeic acid, respectively, were added to 1×10^{-3} mol L⁻¹ solutions of GA and vigorously stirred for 2 min followed by recording of the DPV.

Preparation and determination of real samples

The GA content of the real samples of commercially available orange juice, red and white wine was analysed by using the voltammetric standard addition method. Triplicates of 10 mL of the real samples (orange juice, red and white wine) were used as blanks. Standard addition of aliquots of 1×10^{-2} mol L⁻¹ GA was then added to the electrochemical cell and vigorously stirred using a magnetic stirrer for 2 min, after each addition of the aliquot of GA and the voltammograms recorded.

Results and discussion

Characterisation of the prepared nano-GO-SiO₂ nanoparticles

The nano-GO-SiO₂ nanoparticle-modified electrode was characterised using SEM (scanning electron microscopy). The SEM uses a focused beam with high-energy electrons that generate different signals on the surface of electrode. These signals reveal information about the external morphology, the chemical composition and the visuals of the crystalline structure of the SiO₂ nanoparticles. This shows the spherical morphology and homogenous size distribution (Fig. 1a) of the synthesised SiO₂ nanoparticles. The spherical particles had sizes ranging from 100 to about 200 nm, as can be seen in Fig. 1 a, whilst Fig. 1 b shows the morphology of the nano-GO-SiO₂ nanoparticle mixture on the surface of the GCE.

As reported earlier by Tashkhourian and Nami-Ana [14] and Wang et al. [22], the nanosized SiO₂ nanoparticles as seen on the SEM images creates a large active surface area, better transmission performance and high accumulation efficiency.

Fourier transform infrared spectroscopy (FTIR) was also used for the characterisation of the SiO₂ nanoparticles, nano-GO and nano-GO-SiO₂ nanoparticle combination, to show the decoration of the SiO₂ nanoparticles on the nano-GO. The spectra were recorded as can be seen in Fig. 2, where the nano-GO spectrum and SiO₂ nanoparticles agreed with other works. The bands around 3250 and 1408 cm⁻¹ were attributed to the deformed -OH bonds of graphene oxide and the CO-H groups, respectively. Meanwhile, the bands centred around 1039 cm⁻¹ were associated with the C-O bond stretching and the 1724 cm⁻¹ was then associated to the stretching vibration of the carbonyl or carboxyl groups [23, 24]. With the mixing of the SiO₂ nanoparticles, the characteristic peaks of silica nanospheres were formed on the surface of the nano-

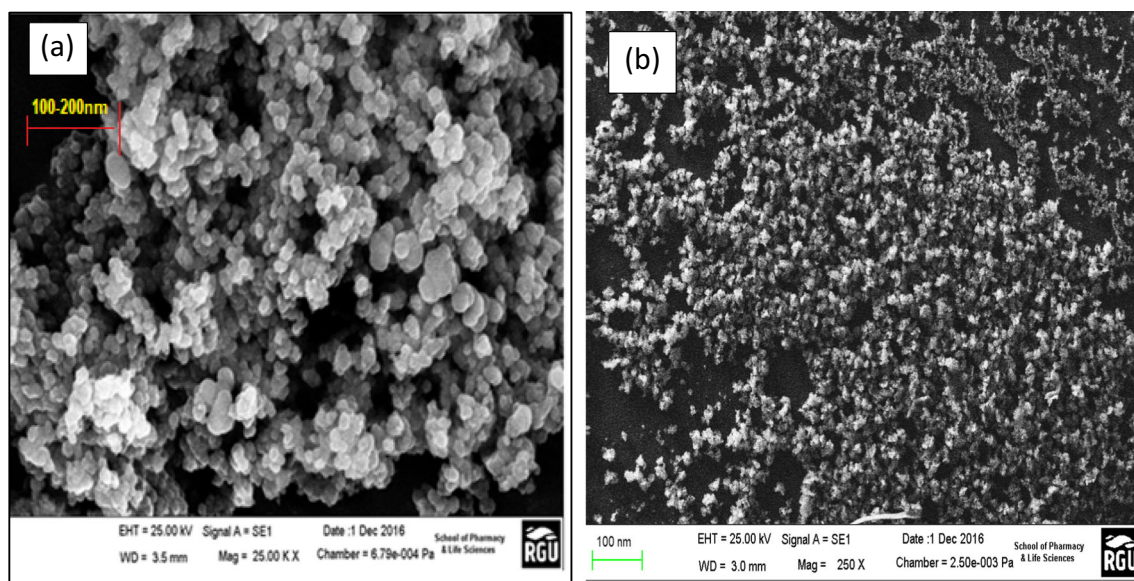


Fig. 1 **a** The SEM image SiO₂ nanoparticles showing the morphology of the nanospheres and the different spectrums used for the EDX, with **b** showing the morphology of the nano-GO-SiO₂ nanoparticles mixture on the GCE

GO, where the peaks at 533 and 795 cm^{-1} are attributed to Si–O–Si bending and symmetric stretching vibration, respectively. The main strong and broad band at 1066 cm^{-1} with a bit of shoulder is usually assigned to the transverse (TO) and longitudinal optical (LO) modes of the Si–O–Si asymmetric stretching vibrations [25, 26]. On the other hand, the band observed at 953.59 cm^{-1} on the SiO₂ IR image attributed to the silanol group (Si–OH) is seen to have diminished at the combination of nano-GO and SiO₂ nanoparticles. This diminished band can be attributed to the hydrogen bonding between the silanol groups (Si–OH) to the GO nanocolloids and the GA. Furthermore, there was no significant change in the area between 1575 and 1727 cm^{-1} of the nano-GO, which would have demonstrated a reaction of the silica with the carbonyl groups forming Si–O–C bonds; hence, a confirmation of the hydrogen bonding proposed mechanism.

The effect and influence of scan rate were studied towards the electrochemical determination of GA, using CV to record the voltammograms of $0.5 \times 10^{-3} \text{ mol L}^{-1}$ of GA in 0.1 mol L^{-1} phosphate buffer of pH 2.0. At increasing scan rates of 10, 25, 50, 75, 100, 150 and 200 mVs^{-1} , the I_p of GA oxidation increased as the scan rate increased as seen in Fig. S1. This shows that the oxidation peak current (I_p) was linear with the scan rate (ν), indicating that the process at the electrode was an adsorption-controlled step. The linear regression equation was $I_p (\mu\text{A}) = 0.25 \pm 0.10, \nu (\text{mVs}^{-1}) + 14 \pm 1$ and $R^2 = 0.9926$.

It can also be observed that, with the increase in the scan rate, there was a positive shift of the peak potentials, which could suggest a kinetic limitation in the reaction [20, 27].

Electrochemical behaviour of gallic acid at the nano-GO-SiO₂ nanoparticle-modified glassy carbon electrode

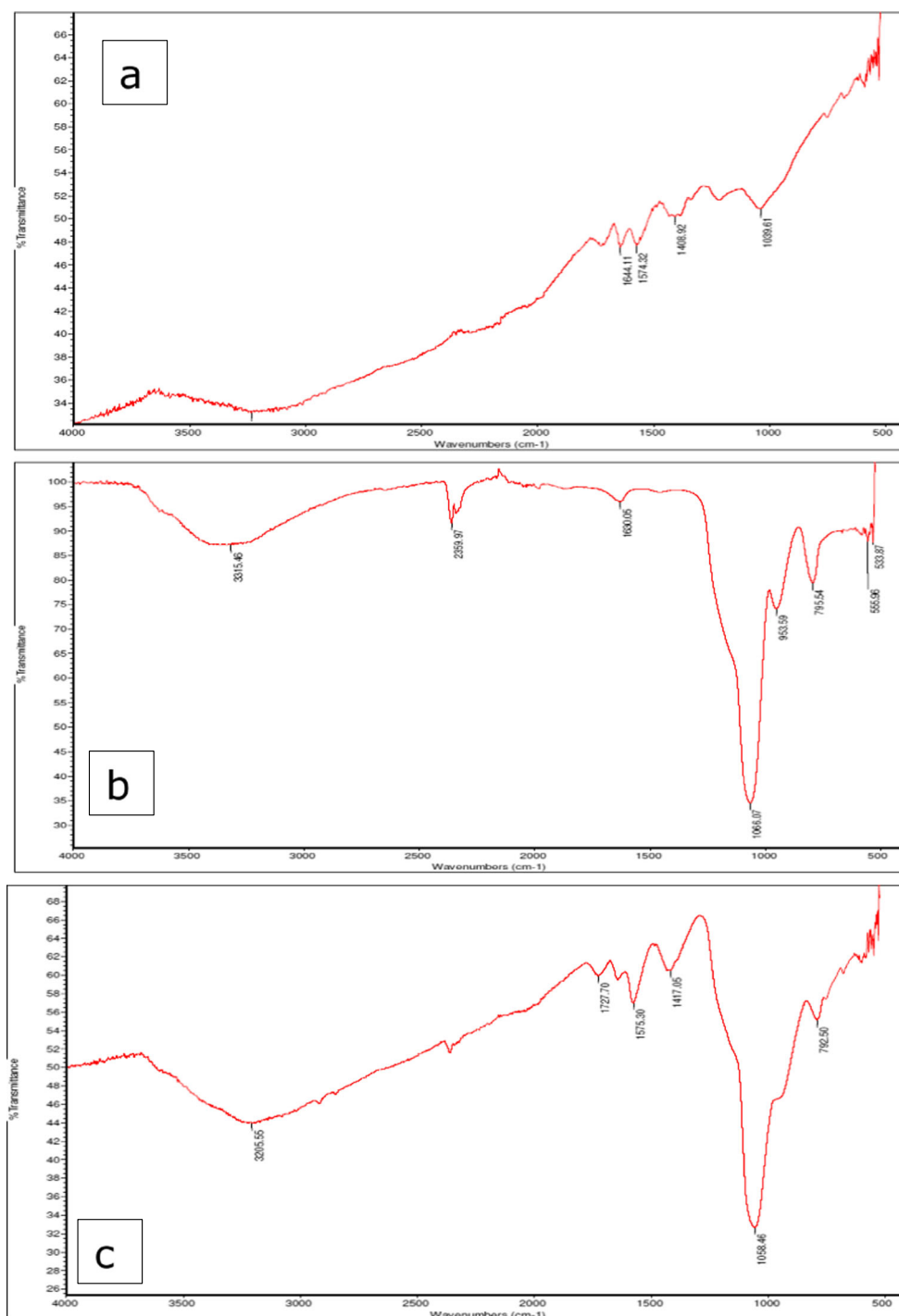
The voltammetric behaviour of gallic acid was studied using cyclic voltammetry (CV). Modification of the GCE electrode by the different reagents was carried out by drop casting with an optimised volume of 5 μL of nano-GO, SiO₂ nanoparticles and nano-GO-SiO₂ nanoparticles. The nano-GO-SiO₂ nanoparticles were dropped on the GCE (see Graphical abstract) and left to dry at room temperature.

The cyclic voltammograms (Fig. 3) show the determination of $1.0 \times 10^{-2} \text{ mol L}^{-1}$ gallic acid using a bare GCE, SiO₂ nanoparticle-modified GCE and nano-GO-SiO₂ nanoparticle-modified GCE (nano-GO-SiO₂-GCE) in phosphate buffer of pH 2 at a scan rate of 100 mVs^{-1} in a potential range of 0.0 to 1.8 V, at room temperature.

The modified electrodes were then used to test GA ($1 \times 10^{-2} \text{ mol L}^{-1}$) and produced peak currents of $261 \pm 1 \mu\text{A}$ at 0.63 V for the SiO₂ nanoparticle GCE, $300 \pm 3 \mu\text{A}$ at 0.63 V for the nano-GO-GCE and $411.7 \pm 0.9 \mu\text{A}$ at 0.64 V for nano-GO-SiO₂ nanoparticles. In contrast, the bare (uncoated) GC electrode produced a peak current (I_p) of $241 \pm 4 \mu\text{A}$ at 0.63 V, which was lower than I_p for the modified electrodes, as can be seen in Fig. 5. Furthermore, no oxidation peak was observed within the potential range of 0.0–1.8 V and specifically, no peaks over the range where GA would normally occur with just the phosphate buffer (0.1 mol L^{-1}) blank.

From the voltammograms shown in Fig. 5, it was deduced that, though the GCE was modified with SiO₂ nanoparticles and nano-GO, the combination of both nano-compounds

Fig. 2 FTIR images showing the spectra of **a** GO nanocolloids, **b** SiO₂ nanoparticles and **c** a combination of the GO nanocolloids and SiO₂ nanoparticles

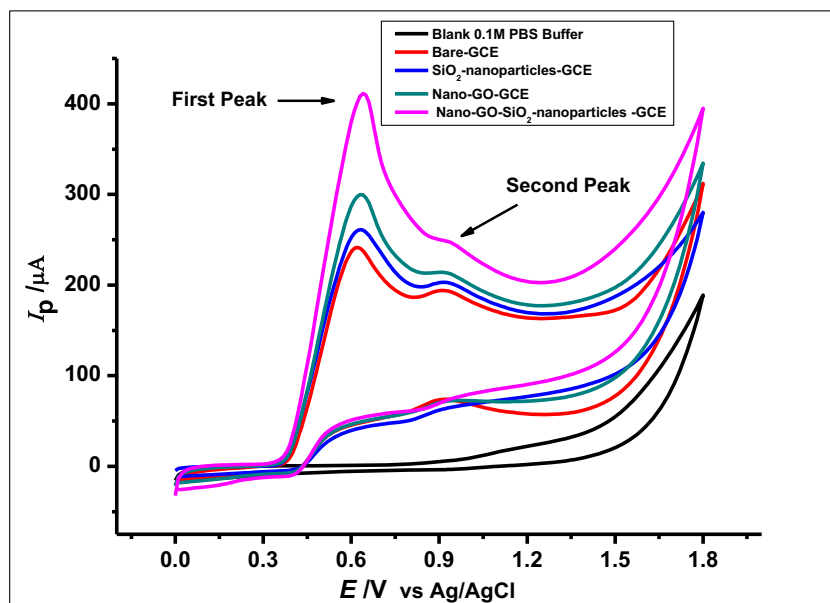


produced better peak currents compared with the individual modifiers.

Gallic acid in this determination showed two oxidation peaks during the anodic sweep between the applied potential ranges of 0.0 to 1.8 V. The two oxidation peaks are found at peak potentials of 0.64 V for the first and 0.91 V for the second peak. These results are similar to other reports in the literature [12, 14, 20].

These two peaks have been characterised in the literature to be the formation of a semiquinone radical (first peak) which is oxidised to a quinone in the second peak [28]. The oxidation that occurs at the first peak is because of the oxidation of the galloyl group, as can be seen in Fig. 4, which then leads to the second peak that is assumed to have been developed from the third –OH group in the galloyl moiety of the compound [14, 28, 29]. This

Fig. 3 Cyclic voltammograms of $1 \times 10^{-2} \text{ mol L}^{-1}$ GA at nano-GO-SiO₂ nanoparticle-modified GCE, SiO₂ nanoparticle-modified GCE, nano-GO-modified GCE and a bare GCE in 0.1 mol L^{-1} phosphate buffer of pH 2.0 at a scan rate of 100 mVs^{-1} (S/N = 3)



assumption is reinforced by the fact that the electro-oxidation of the carboxylic group ($-\text{COOH}$) occurs at 2.0 V and produces CO_2 [12].

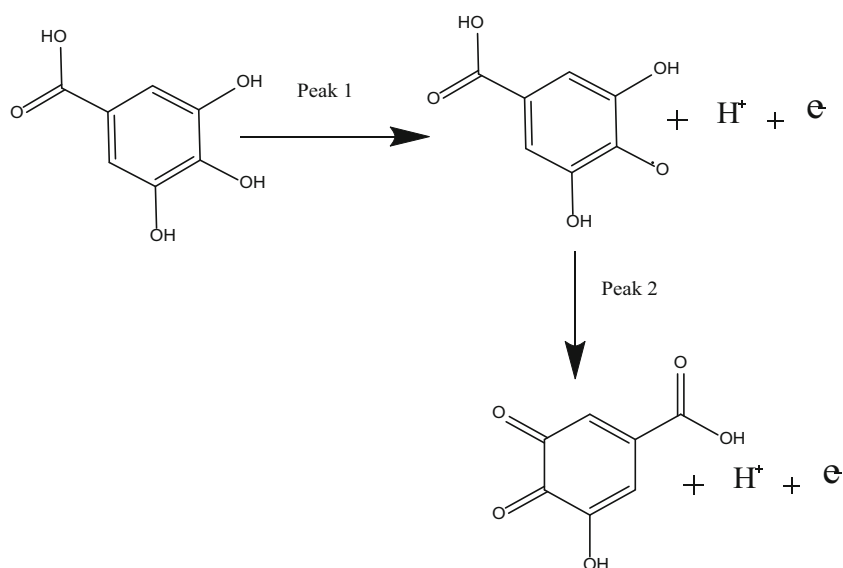
Suggested mechanism of action

The chemical effect of nano-GO-SiO₂ nanoparticles towards the oxidation of GA and the enhancement of the voltammograms can be attributed to the structural, chemical and physical properties of GO and SiO₂. The SiO₂ nanoparticles with silanol groups bind with the GO nanocolloids via hydrogen bonding as depicted in Fig. 5. The presence of hydrogen bonds on the surface of SiO₂ under aqueous conditions,

contributes to the electrical transport properties of graphene oxide on SiO₂ surface structures. Whilst adsorbed on the surface of the graphene oxide nanocolloids, SiO₂ nanoparticles also adsorb with GA.

The mechanism proposed for this work is based on the work of S. Milonjić [30, 31] and colleagues, where they investigated the interaction of SiO₂ and GA. They found out that SiO₂ and GA underwent an adsorption interaction which was linked to the pK_a (4.41) of GA, where the amount of adsorbed GA increases with increasing equilibrium concentration of GA in the solution or decreasing pH of the solution. Meanwhile, there was a decrease in the amount adsorbed when there was an increase in pH of the solution.

Fig. 4 The reaction mechanism of oxidation of gallic acid, showing the two peaks in CV and DPV in scheme 1



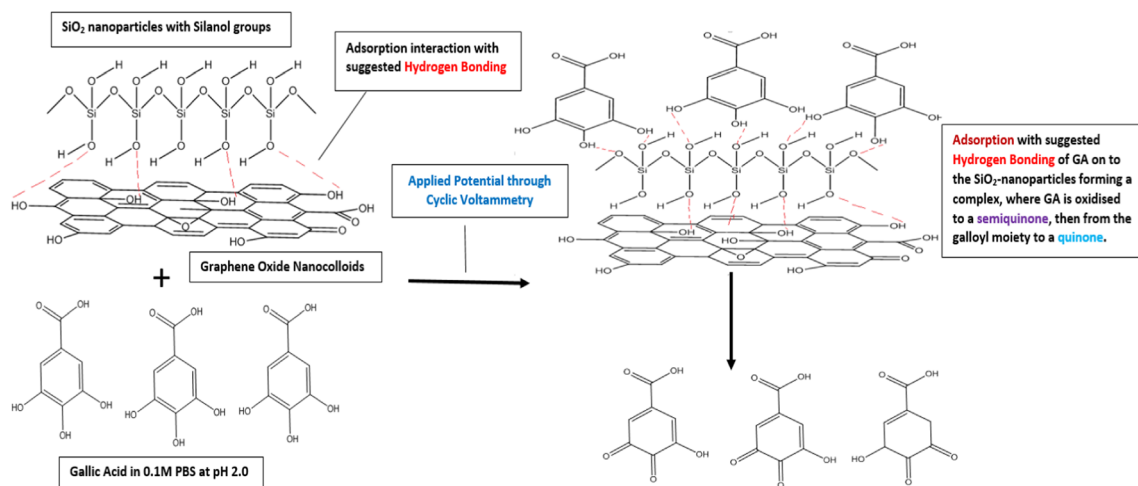


Fig. 5 Scheme showing the interaction of SiO_2 nanoparticles and graphene oxide nanocolloids as they interact with gallic acid by adsorption with a suggestion of hydrogen bonding, as applied potential is sent through the modified electrode

The chemical behaviour of SiO_2 and GA was then explained using the pK_a of GA molecules and the surface charge of the silica powder. It should be noted that the adsorption of weak organic electrolytes like GA on the surface of amphoteric solids like silica depends amongst other things on the degree of dissociation of the electrolyte and the surface charge of the solid, which all depends on the pH of the aqueous solution [31]. The point of zero charge values (pH_{pzc}), for various forms of SiO_2 , lie between the pH range of 2.5–3.6 [30, 31]; hence, when the pH is below the pH_{pzc} , the surface of SiO_2 becomes positively charged, whilst in a pH higher than the pH_{pzc} , the surface becomes negatively charged. With a pK_a of a weak acid, GA at pH values less than 3.5 is mostly undissociated or in its stable form and dissociates when the pH increases.

It can therefore be interpreted that the maximum adsorption of GA to SiO_2 occurs at a pH_{pzc} where the silica surface is uncharged, and the GA molecule is un-dissociated. Hence, in a phosphate buffer solution of $\text{pH } 2.0 < 3.5$, where the electrochemical determination takes place, GA is adsorbed on the SiO_2 . Meanwhile, at $\text{pH} > 3.5$, GA's dissociation increases, leading to an increase in the negative ion $\text{C}_6\text{H}_2(\text{OH})_3\text{COO}^-$ concentration and a negatively charged SiO_2 surface. With the increase in the negative molecules and negative surface, the adsorption between GA and SiO_2 nanoparticles decreases due to the repulsive negative-negative forces in the solution.

On the other hand, the GO nanocolloids used possess excellent electrochemical properties due to their favourable electron mobility and unique surface properties like high specific surface area that would accommodate active species like GA and facilitate electron transfer [32]. The capacity of GO to be chemically or physically functionalised, showing non-covalent π - π (π -stacking), cation- π , van der Waals or hydrogen bonding [33], formed the bases of this proposed

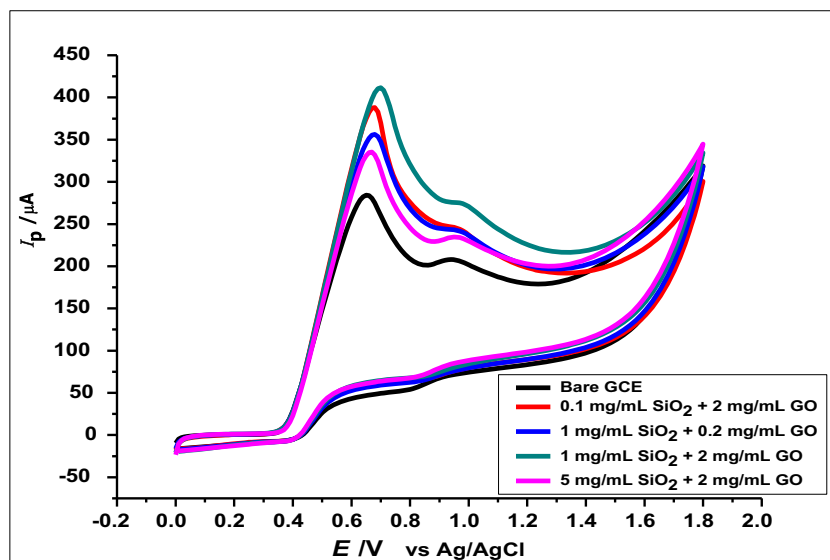
mechanism. The SiO_2 nanoparticles physically functionalise the GO nanocolloids, thus enhancing the availability of the nanosized surface area for electron transfer, whilst the adsorption of the GA to SiO_2 and nanosized area provides better mass transport of the GA to the electroactive sites on the electrode surface. Meanwhile, Zhang and Choi [23] proposed an electrorheological characteristic of Si-GO hybrid composite, hence easy transportation of electrons. Moreover, the conductive GO nanocolloids support facilitates the efficient collection and transfer of electrons to the GCE surface as described by Chen and colleagues [34].

Furthermore, an alternative and plausible explanation for the phenomenon observed in the synergistic effect of the nano-GO and SiO_2 nanoparticles could be that of voltammetry at a three-phase junction, discussed by Oldham and Scholz et al. in their work [35–38]. They asserted that when insoluble insulating crystals like SiO_2 nanoparticles adhere to an electrode, the three-phase junction (where the electrolyte solution, electrode and crystal meet) is the only feasible site for an electrochemical reaction to occur. Therefore, this implies that the redox process of gallic acid might have occurred at the three-phase junction between graphene oxide nanocolloids and SiO_2 nanoparticles.

Effect of concentration changes of modifier

The effect of the changes in concentration of the nano-GO and the SiO_2 nanoparticles in the modifying mixture was investigated. The following concentrations (0.1 mg/mL SiO_2 nanoparticles + 2 mg/mL nano-GO; 1 mg/mL SiO_2 nanoparticles + 0.2 mg/mL GO; 1 mg/mL SiO_2 nanoparticles + 2 mg/mL GO and 5 mg/mL SiO_2 nanoparticles + 2 mg/mL GO) were used for the modification of the glassy carbon electrode and used to determine $1 \times 10^{-2} \text{ mol L}^{-1}$ GA. The combinations studied

Fig. 6 Voltammograms showing changes in the constitution of the nano-GO-SiO₂ nanoparticles used to determine 1×10^{-2} mol L⁻¹ GA in 0.1 mol L⁻¹ phosphate buffer at pH 2.0 using CV at a scan rate of 100 mVs⁻¹



had equal amounts of nano-GO and SiO₂ nanoparticles of variable concentration, sonicated for 1 h to enable uniform dispersion and bonding of the two compounds. From the results (Fig. 6), the combination of 1 mg/mL SiO₂ nanoparticles + 2 mg/mL GO nanocolloids showed the highest peak current (I_p) of 411.7 μ A; hence, this combination was subsequently used for modification of the GCE.

Effect of changes in the volume of modifier on GCE

The quantity of nano-GO-SiO₂ nanoparticle drop-cast on the glassy carbon electrode and the effect on the determination of 1×10^{-2} mol L⁻¹ GA in a 0.1 mol L⁻¹ phosphate buffer at a pH of 2.0, using CV at a scan rate of 100 mVs⁻¹, were investigated. Volumes of 2 to 5 μ L were studied, and it was observed

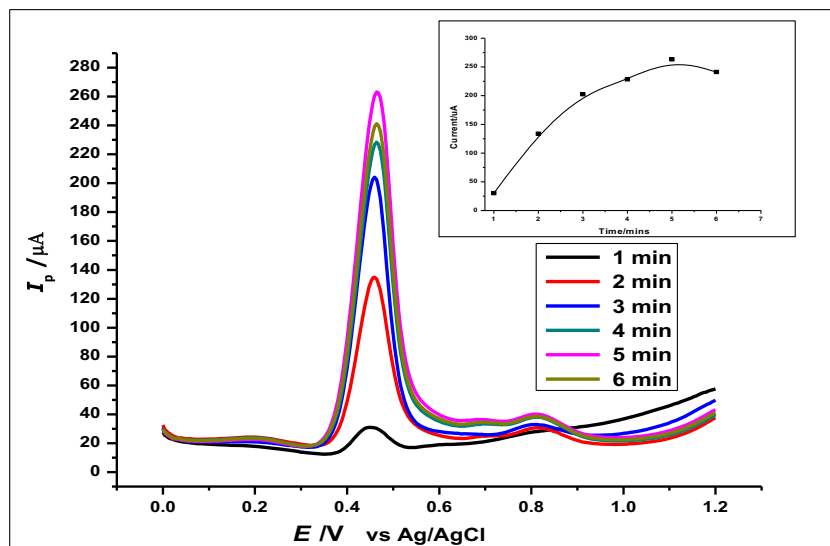
that 5 μ L of nano-GO-SiO₂ nanoparticles gave the highest peak current (I_p) of 405.18 μ A, as a further increase in the volume made no difference, but instead reduced the peak current, as they seem to foul the electrode surface. The volume of the mixture used for the GCE modification for further experiments was 5 μ L.

Effect of adsorption time

The effect of the adsorption of GA to the electrode with time was demonstrated by the detection of GA (1×10^{-3} mol L⁻¹) in a 0.1 mol L⁻¹ phosphate buffer of pH 2.0 at room temperature using DPV at a scan rate of 100 mV/s⁻¹.

The voltammograms in Fig. 7 demonstrated the adsorption of gallic acid onto the nano-GO-SiO₂ nanoparticle electrode

Fig. 7 DPV voltammograms showing the effect of adsorption time in the determination of 1×10^{-3} mol L⁻¹ GA in 0.1 mol L⁻¹ phosphate buffer of pH 2.0 at a scan rate of 100 mVs⁻¹ (inset) and the plots showing the same effect of adsorption time



by showing a sharp increase after the first scan to an optimum by the fourth to fifth minute, reaching saturation of the GA on the electrode after 6 min. This phenomenon confirms the proposed interaction mechanism of GA and the nano-GO-SiO₂ nanoparticles as being an adsorption reaction and is in agreement with the work of Tashkhourian [14].

Effect of pH on gallic acid oxidation

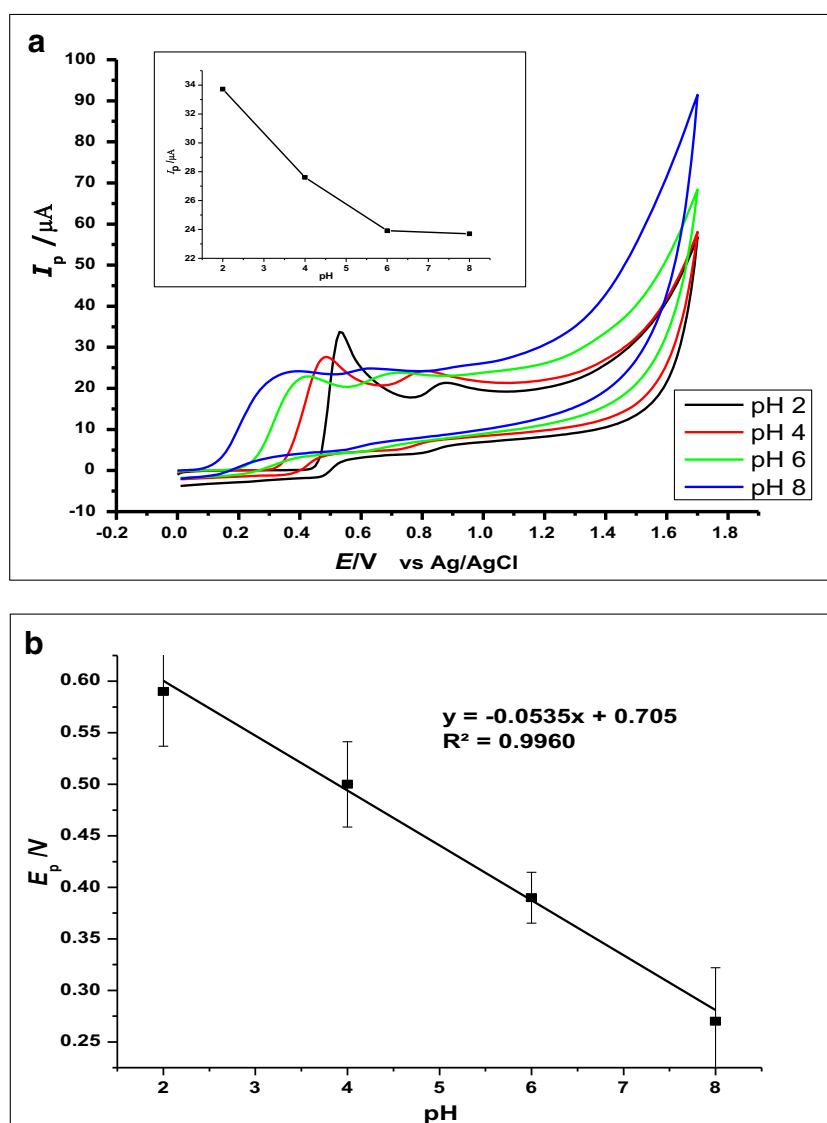
The effect of the pH of the gallic acid solution towards its electrochemical activity and the activity of the nano-GO-SiO₂-GCE were investigated. The electrochemical response of gallic acid (1×10^{-4} mol L⁻¹) was studied with pH ranging from 2.0 to 8.0 using cyclic voltammetry at a scan rate of 100 mVs⁻¹, in phosphate buffer (0.1 mol L⁻¹). The oxidation peak and the peak potential (E_p) shown have clearly been influenced by the pH, as can be seen in Fig. 8. From the

voltammograms, a pH value of 2.0 showed a well-shaped oxidation peak, with the highest peak current (I_p) of 33.72 μ A.

On the other hand, as the pH increases, there was a reduction in the peak current (Fig. 8a). For example, at pH 8, the lowest peak current of 23.70 μ A was observed. There was also a negative shift of the oxidation peak potential from 0.56 V downwards to 0.31 V, as can be seen in Fig. 8a, with a gradual broadening of the oxidation peak/peak area with the increase in pH.

The relationship between the pH and the peak potential (E_p) shows a good linear relationship as can be seen in Fig. 8b in the pH range of 2.0 to 8.0, with a linear regression equation of $E_{pa} \text{ V} = 0.70 \pm 0.01 - 0.05 \pm 0.03 \text{ pH}$, with $R^2 = 0.9960$. The slope of the regression line, 54 mV/pH, is comparable with the Nernstian value of 59 mV/pH at 25 °C, for equal number of proton and electron transfer reactions. The pH and peak potential relationship is thus consistent with a

Fig. 8 **a** Cyclic voltammograms showing the effect of pH on the electrochemical behaviour of 1×10^{-4} mol L⁻¹ gallic acid (inset). Plot of the peak current (I_p) against different pH values showing the effect of pH on the electrochemical behaviour of 1×10^{-4} mol L⁻¹ gallic acid using bare glassy carbon electrode at a scan rate of 100 mVs⁻¹. **b** Plot of the peak potential E_p on detection of 1×10^{-4} mol L⁻¹ gallic acid using nano-GO-SiO₂-GCE as a function of pH, measured at a scan rate of 100 mVs⁻¹



two-proton/two-electron reaction, which is consistent with other literature data [14, 20]. Based on the results, the pH condition selected as the most appropriate for the oxidation of gallic acid in this experiment was pH 2.0.

This may be attributed to the fact that GA, with a pK_a of 4.4 being a weak organic acid, is most stable or is in its non-dissociated state at pH 2.0. This pH was the most appropriate for the adsorption of GA on to SiO_2 .

Limit of detection for the electrochemical oxidation of GA

Differential pulse voltammetry was used for the investigation of different concentrations of GA, to determine the limit of detection (LOD) at a scan rate of 100 mVs^{-1} and pulse amplitude of 80 mV. With the increase in concentration studied in the range of 6.25×10^{-6} to $1.0 \times 10^{-3} \text{ mol L}^{-1}$, there was a correlated increase in the peak current, whilst using the nano-GO- SiO_2 nanoparticle-modified electrode. The measurements produced voltammograms (Fig. 9) in which the peak current increased proportionally with increasing gallic acid concentration. Using DPV in this measurement produced two oxidation peaks at peak potentials of 0.45 V and 0.8 V as different concentrations of GA were being determined.

Using the first main GA oxidation peak, the analytical calibration graph as seen in Fig. 9 (inset) showed a linear relationship between the peak current and the increase in gallic acid concentration, producing a linear regression equation, where $I_p (\mu\text{A}) = 221 \pm 5.5, C (\text{mmol L}^{-1}) + 9.43 \pm 2$ and $R^2 = 0.9956$ within a range of 6.25×10^{-6} to $1.0 \times 10^{-3} \text{ mol L}^{-1}$. The limit of detection (LOD), defined as $(3 \times \text{Std}_{\text{Blank}})/m$,

where $\text{Std}_{\text{Blank}}$ is the standard deviation of the blank and m is the slope, was found to be $2.09 \times 10^{-6} \text{ mol L}^{-1}$.

The proposed method is comparable with other methods based on the maximum permitted antioxidant levels in food within EU and North American guidelines, which ranges from 20 to 1000 ppm (20 to 1000 mg L^{-1}) [39]. Hence, for GA, the permitted range would be $1.2 \times 10^{-1} \text{ mol L}^{-1}$ to 5.9 mol L^{-1} . Thus, with an LOD of $2.09 \times 10^{-6} \text{ mol L}^{-1}$ which is far below these standard permitted levels, the electrode is comparable with others, as can be seen in Table 1.

Reproducibility and repeatability of the method

The reproducibility of the nano-GO- SiO_2 -modified glassy carbon electrode was investigated, by using the modified electrode to determine the oxidation peak current produced by $1.0 \times 10^{-3} \text{ mol L}^{-1}$ GA. The oxidation peak currents produced by eight replicates of $1.0 \times 10^{-3} \text{ mol L}^{-1}$ GA independently measured with eight different electrodes, with a relative standard deviation of 3.8% indicated a good reproducibility.

Meanwhile, the repeatability of the method was investigated by using a nano-GO- SiO_2 -modified glassy carbon electrode, for eight repetitive determination of $1.0 \times 10^{-3} \text{ mol L}^{-1}$ gallic acid. The relative standard deviation of the oxidation peak current was found to be 2.92%, which shows good repeatability of the method.

Stability of electrode

The stability of the nano-GO- SiO_2 nanoparticle-modified glassy carbon electrode was also investigated. The electrode

Fig. 9 Differential pulse voltammograms of various concentration of GA at nano-GO- SiO_2 nanoparticles in 0.1 mol L^{-1} phosphate buffer at pH 2.0 at a scan rate of 100 mVs^{-1} with voltammograms (a–j) that corresponds to the following concentrations: (a) 0.0, (b) $0.00625 \text{ mmol L}^{-1}$, (c) $0.0125 \text{ mmol L}^{-1}$, (d) $0.025 \text{ mmol L}^{-1}$, (e) 0.05 mmol L^{-1} , (f) $0.075 \text{ mmol L}^{-1}$, (g) 0.1 mmol L^{-1} , (h) 0.25 mmol L^{-1} , (i) 0.5 mmol L^{-1} , (j) 0.75 mmol L^{-1} , (k) 1 mmol L^{-1} . (Inset) The calibration curve for the determination of GA

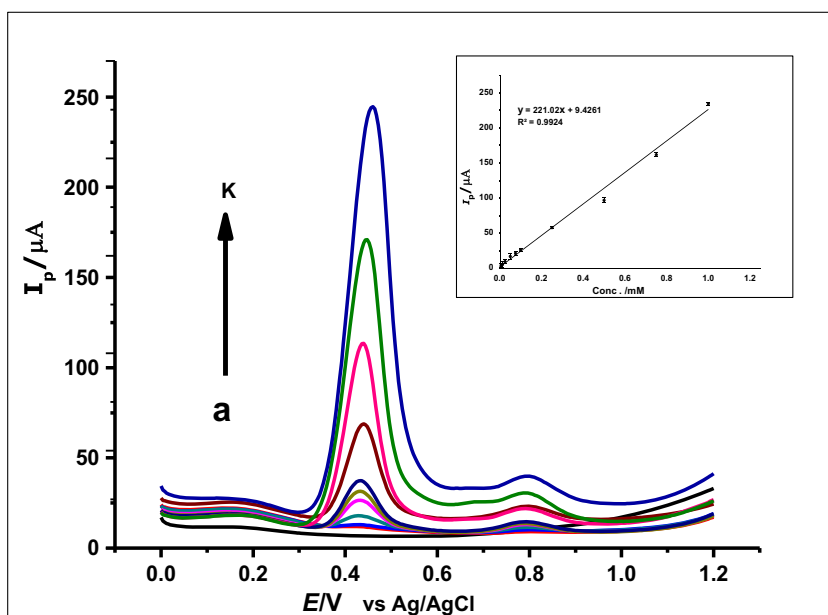


Table 1 Some electrochemical methods used in the determination of gallic acid and the efficiency of such methods

Electrode	Method	Medium analysed	Linear range	Limit of detection	Reference
Graphite electrode modified with thionine (TH) and nickel hexacyanoferrate (NiHCF)	Cyclic voltammetry, differential pulse voltammetry	Green tea, milk	4.99×10^{-6} to 1.2×10^{-3} M	1.66×10^{-6} M	[30]
Carbon paste electrode modified with carbon nanotubes	Cyclic voltammetry, differential pulse voltammetry	Red wine, white wine	5×10^{-7} to 1.5×10^{-5} M	3.0×10^{-7} M	[31]
Carbon paste electrode modified with TiO ₂ nanoparticles	Differential pulse voltammetry	Tap water, green tea, black tea	2.5×10^{-6} to 1.5×10^{-4} M	9.4×10^{-7} M	[15]
Carbon paste electrode modified with SiO ₂ nanoparticles	Differential pulse voltammetry	Tap water, orange juice, green tea, black tea	8.0×10^{-7} to 1.0×10^{-4} M	2.5×10^{-7} M	[14]
Glassy carbon electrode modified with the electrodeposition of Zn-Al-NO ₃ layered double hydroxide film.	Differential pulse voltammetry	Green tea	4.0×10^{-6} to 6×10^{-4} M	1.6×10^{-6} M	[22]
Glassy carbon electrode modified with polyethyleneimine-functionalised graphene oxide (PEI-rGO)	Cyclic voltammetry	Green tea, black tea	5.8×10^{-7} to 5.8×10^{-4} M	4.1×10^{-7} M	[23]
Glassy carbon electrode modified with nano-GO-SiO ₂ -nanoparticles	Cyclic voltammetry	Red wine, white wine, orange juice	6.2×10^{-6} to 1.0×10^{-3} M	2.09×10^{-6} M	This work

was kept for 30 days and was used for the determination of 1.0×10^{-3} mol L⁻¹ gallic acid and the oxidation peak currents produced from the readings of the first day and that of the 30th day showed an RSD of 6.58%, depicting a relatively good stability.

Selectivity of the method

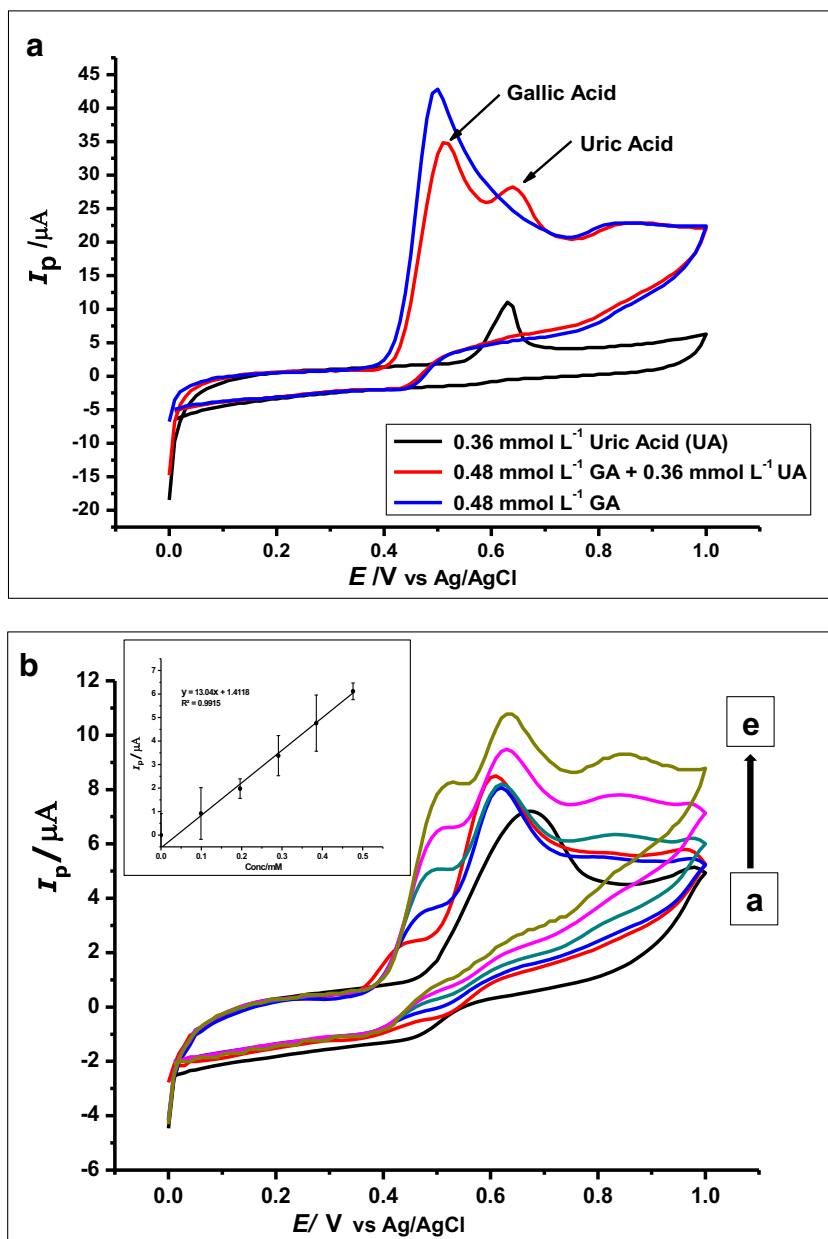
The selectivity of the modified glassy carbon electrode was tested, by the simultaneous detection of GA and uric acid (UA) in a 0.1 mol L⁻¹ phosphate buffer solution at a pH of 2.0 using CV at a scan rate of 100 mVs⁻¹, at room temperature. This was done by the addition of 0.48 mmol L⁻¹ of GA into a 10-mL cell of 0.36 mmol L⁻¹ uric acid and voltammograms recorded, as seen in Fig. 10 a. A concentration of 0.48 mmol L⁻¹ of GA was used, because it was at this concentration that a significant distinction of the voltammograms of GA and UA was observed. GA and UA showed distinct peaks at approximately 0.51 V and 0.64 V, respectively, with the smaller second GA oxidation at a peak potential of 0.88 V. Meanwhile, the voltammograms of UA on its own without GA showed an oxidation peak at 0.64 V and that of GA on its own show oxidation peaks at 0.5 V for the first peak and at 0.88 V for the second smaller peak (Fig. 10b).

To confirm the selectivity of the electrode, the UA concentration was kept constant at 0.36 mmol L⁻¹ in a 10-mL voltammetric cell and then aliquots of 100 µL of 10 mmol L⁻¹ GA were intermittently added and then vigorously stirred for 2 min. Voltammograms were then recorded, as can be seen in Fig. 10 b. In this case, the nano-GO-SiO₂ nanoparticle-modified GCE produced the voltammograms as can be seen in Fig. 10 b, where increasing the concentration of GA showed a proportional increase in the peak current. Hence, Fig. 10 b (inset) shows a linear relationship between the peak current and the increase in gallic acid concentration, producing a linear regression equation where I_p (µA) = 13.0 ± 0.6 , C (mmol L⁻¹) + 1.41 ± 0.20 and $R^2 = 0.9915$ within a range of 0.1 to 0.48 mmol L⁻¹.

Interferences

The interference of various species in the determination of 1×10^{-3} mol L⁻¹ gallic acid was also investigated (Table 2). This was carried out by adding different amounts of foreign ions to a known quantity of the analyte as it was being determined. The foreign species that were used in this case for the investigation of interference were K⁺, Ca²⁺, Fe³⁺ and Na⁺, then ascorbic acid, caffeine, caffeic acid and quercetin, respectively. The selection of some of the cations was based on reports of their complexation with GA in the literature [40, 41] and the

Fig. 10 **a** Cyclic voltammograms showing 0.36 mmol L⁻¹ uric acid concentration, 0.48 mmol L⁻¹ GA and a mixture of the two concentrations of GA and UA in a 0.1 mol L⁻¹ phosphate buffer solution at a pH 2.0 at scan rate 100 mVs⁻¹. **b** Cyclic voltammograms showing the constant 0.36 mmol L UA concentration and increasing GA concentration from 0.1 to 0.48 mmol L⁻¹ in 0.1 mol L⁻¹ phosphate buffer at pH 2.0 at a scan rate of 100 mVs⁻¹. (Inset) A graph showing the increasing concentration of GA against the increasing peak current in a constant UA concentration of 3.6×10^{-4} mol L⁻¹

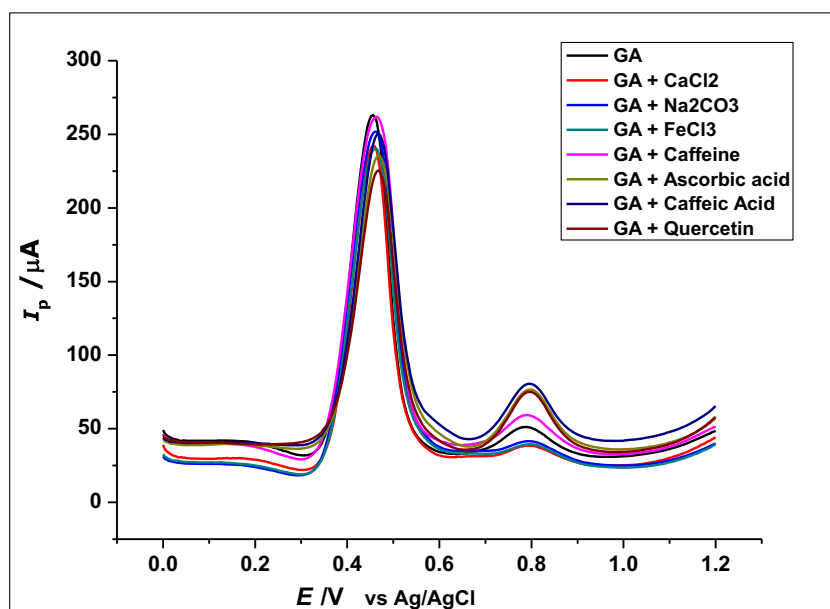


possibility of these compounds interfering with the electrochemical determination of GA. The tolerable limits of these ions for interference was defined as the highest amounts of foreign ions that would produce an error of not more than 5% in the determination of the analyte, which in this case is gallic acid. With relative standard error values of less than 5%, these ions did not interfere with the determination of gallic acid at the first GA oxidation peak, as can be seen in the results shown in Fig. 11. However, there were some minor changes in the second GA oxidation peak, which is not normally the peak used for electrochemical determination of GA.

Table 2 Effects of various substances on the determination of 1×10^{-3} mol L⁻¹ gallic acid

Interfering species	Amount in solution (mol L ⁻¹)	Relative standard deviation (%)
Ca ²⁺	1×10^{-1}	± 3.70
Na ⁺	1×10^{-1}	± 1.29
Fe ³⁺	1×10^{-1}	± 1.93
Cl ⁻	1×10^{-1}	± 2.01
CO ₃ ²⁻	1×10^{-1}	± 1.45
Ascorbic acid	1×10^{-3}	± 0.15
Caffeic acid	1×10^{-3}	± 2.80
Caffeine	1×10^{-3}	± 3.89
Quercetin	1×10^{-3}	± 4.64

Fig. 11 DPV voltammograms of $1 \times 10^{-3} \text{ mol L}^{-1}$ gallic acid with different concentrations of some known interfering compounds in 0.1 mol L^{-1} phosphate buffer at pH 2.0 and a scan rate of 100 mVs^{-1}



Analytical application of the method

This analytical method with the nano-GO-SiO₂ nanoparticle-modified electrode was used for the determination of GA in red wine, white wine and orange juice. The real samples (i.e. red wine, white wine and orange juice) were used based on previous work from other group showing the presence of GA in those types of samples [14]. However, in red wine, white wine and orange juice GA were not detected. This may be due to the origin and nature of the samples. Hence, spiking methodology was applied to all samples to clearly demonstrate the potential of our proposed sensor as shown in Table 3. The experiments were done by using 10 mL of the real samples as blanks and then the standard addition of known concentrations of aliquots of gallic acid within a concentration range of 9.0×10^{-4} to $4.7 \times 10^{-3} \text{ mol L}^{-1}$ was added to the cells containing the real samples and the voltammograms recorded and the results calculated as can be seen in Table 3.

Conclusion

In this paper, for the first time, graphene oxide nanocolloids mixed with SiO₂ nanoparticles synthesised by the sol-gel method were drop-cast on a glassy carbon electrode. This electrode was then used for the electrochemical determination of gallic acid in a 0.1 mol L^{-1} phosphate buffer at pH 2.0. Cyclic and differential pulse voltammetry were used as the electrochemical techniques and it was found that graphene nanocolloids mixed with SiO₂ nanoparticles showed an electrochemical effect towards the oxidative determination of gallic acid, within a concentration range of 6.2×10^{-6} to $1.0 \times 10^{-3} \text{ mol L}^{-1}$.

The modified electrode demonstrated selectivity towards the determination of gallic acid and uric acid, which is a marked advantage over other electrodes in the literature.

The electrode was found to be a simple, sensitive, fast and cost-effective method for the determination of gallic acid; it

Table 3 Results of the analysis of GA in spiked red wine, white wine and orange juice and the validation HPLC result of red wine

Samples	Added (mol L^{-1})	Found (mol L^{-1})	Relative error	Recovery (%)
Red Wine	0.0	Undetected	—	—
	1.66×10^{-4}	1.689×10^{-4}	± 2.32	102.3
	2.30×10^{-4}	2.47×10^{-4}	± 7.39	107.4
White Wine	0.0	Undetected	—	—
	1.66×10^{-4}	1.583×10^{-4}	± 4.64	95.4
	2.30×10^{-4}	2.42×10^{-4}	± 5.65	105.7
Orange Juice	0.0	Undetected	—	—
	1.70×10^{-4}	1.66×10^{-4}	± 2.36	97.6
	2.30×10^{-4}	2.379×10^{-4}	± 3.44	103.4

showed very good reproducibility, repeatability and a relatively good long-term stability.

The electrochemical sensor was successfully applied as a sensitive electrochemical method for the determination of gallic acid in red wine, white wine and orange juice within the concentration range of 9.0×10^{-4} to 4.7×10^{-3} mol L⁻¹. The results showed the present method to be comparable with some of the other methods in the literature, as it detects far below the maximum permitted gallic acid levels in food within the EU and North America.

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